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(57) Abstract

The invention provides isolated nucleic acid compounds, proteins and peptides, said proteins and peptides being related to the VEGF family of angiogenic ligands. Also provided are vectors and transformed heterologous host cells for expressing the proteins. Also provided are methods for identifying compounds that bind and/or modulate the activity of said proteins, methods for treating cancer or inhibiting tumor growth by administration of an antagonist of VEGF-R, and methods for stimulating angiogenesis in a patient in need thereof by administration of a therapeutically effective amount of VEGF-R.

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VEGF RELATED GENE AND PROTEIN

5 BACKGROUND OF THE INVENTION

This invention relates to recombinant DNA technology. In particular the invention pertains to a vascular endothelial growth factor (VEGF-R)gene, and its encoded protein. Also contemplated are methods for identifying compounds that bind said protein, and/or antagonize said protein, and methods for stimulating angiogenesis, as well as methods for inhibiting tumor growth in mammals by administration of an antagonist of said VEGF-R protein.

The growth of solid tumors and the formation of metastases are dependent on the formation of new blood vessels. Among the various angiogenic factors, VEGF plays a pivotal role in tumor angiogenesis. VEGF is expressed and secreted by most solid tumors. Moreover, VEGF is a specific mitogen for endothelial cells that induces endothelial cell migration, formation of tube-like structures, and vascular permeability. In addition, VEGF receptors are expressed preferentially in vessels lining and penetrating tumors. Thus, the family of VEGF molecules are of therapeutic interest as targets for inhibiting tumor growth. Moreover, VEGF molecules are of therapeutic interest in promoting angiogenesis, for example, as a treatment for cardiac artery blockage or for the treatment of atherosclerosis.

VEGF is a heat-stable, 46 KD dimeric protein with structural similarity to placental growth factor (PIGF) and

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more distant homology to the platelet-derived grwoth factors. Ferrara, N. et.al. Endocr. Rev. 13, 18-32, 1992. The biological activites of VEGF are mediated by two transmembrane receptor tyrosine kinases, termed flt-1 and flk-1/KDR, that are expressed predominantly on vascular endothelial cells and their embryonic progenitors.

The biological activity of VEGF and cognate molecules make them prime candidates for therapeutic development. For example, injections of VEGF protein, and DNA molecules encoding VEGF, have resulted in formation of new vessels in embryonic tissues. Drake CJ and Little CD. Proc. Nat. Acad. Sci. 92, 7657-661, 1995. VEGF administration may also be useful for angiogenesis in vivo in adult tissues through local administration to the target tissue.

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BRIEF SUMMARY OF THE INVENTION

The present invention provides isolated nucleic acid molecules that encode novel molecules related to the VEGF family, termed herein "VEGF-R". The progenitor molecule disclosed herein was identified from a human source. Having 20 the cloned VEGF-R gene enables the production of recombinant VEGF-R protein, the isolation of orthologous genes from other organisms, and/or paralogous genes from the same organism, chromosome mapping studies, and the implementation of large scale screens to identify compounds that inhibit 25 the binding of said VEGF-R or related molecule to its cognate receptor protein, as a means to identify potential pharmaceutical compounds useful for inhibiting tumor growth or for treating cancer. The proteins, peptides, and analogs thereof, described herein are also useful therapeutic

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agents, for stimulating angiogenesis in a patient in need thereof, for example, in the treatment of atherosclerosis, or cardiopathies involving blood vessel blockage, or diminished blood flow.

In one embodiment the present invention relates to VEGF-R protein (SEQ ID NO:2), and analogs thereof that are structurally and/or functionally related to SEQ ID NO:2.

In another embodiment the present invention relates to a modified, soluble VEGF-R protein comprising a carboxyl end fragment of SEQ ID NO:2.

In another embodiment the present invention relates to an isolated nucleic acid molecule encoding VEGF-R protein, and related proteins described herein.

In another embodiment, the invention relates to a nucleic acid molecule comprising the nucleotide sequence identified as SEQ ID NO:1, and/or residues 276 through 1310 of SEQ ID NO:1.

In another embodiment, the present invention relates to a nucleic acid that encodes SEQ ID NO:2, or functional fragment thereof.

In another embodiment, the present invention relates to a nucleic acid that hybridizes to a nucleic acid defined by residues 276 through 1310 of SEQ ID NO:1 under high stringency conditions, and encodes a protein that is capable of inducing angiogenesis, and/or in treating blood vessel blockage in vivo or in vitro.

In yet another embodiment, the present invention relates to a recombinant DNA vector that incorporates the VEGF-R gene (residues 276 through 1310 of SEQ ID NO:1) in

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operable-linkage to gene expression sequences, enabling said gene to be transcribed and translated in a host cell.

In still another embodiment the present invention relates to host cells that have been transformed or transfected with the cloned VEGF-R gene such that said gene is expressed in the host cell.

In another embodiment, the present invention relates—to—a method for treating cancer, and/or in preventing or inhibiting tumor growth, and/or in causing shrinkage of a cancerous tumor, in vivo or in vitro by treatment with an antagonist of VEGF-R.

In yet another embodiment, the present invention relates to a pharmaceutical formulation comprising as an active ingredient a therapeutically effective amount of an VEGF-R, associated with one or more pharmaceutically acceptable carriers, excipients, or diluents thereof.

In a further embodiment, the present invention relates to a pharmaceutical formulation comprising an VEGF-R antagonist associated with one or more pharmaceutically acceptable carriers, excipients, or diluents thereof.

In still another embodiment the present invention relates to a method for identifying an antagonist of an VEGF-R, wherein said antagonist interferes in the binding of said VEGF-R to its receptor, comprising the steps of: mixing an VEGF-R protein preparation with a test sample, or a control sample; monitoring the binding of said VEGF-R protein to its receptor, by any suitable means; and comparing the level of binding of VEGF-R to its receptor in the test sample to the level of said binding in the control sample.

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DETAILED DESCRIPTION OF THE INVENTION Definitions

The term "analog" or "functional analog" refers to a modified form of VEGF-R in which at least one amino acid substitution has been made such that said analog retains substantially the same biological activity as the unmodified VEGF-R in vivo and/or in vitro.

"Angiogenesis" refers to the phenomenon of blood vessel formation and differentiation in vivo and/or in vitro.

The terms "complementary" or "complementarity" as used herein refer to the capacity of purine and pyrimidine nucleotides to associate through hydrogen bonding to form double stranded nucleic acid molecules. The following base pairs are related by complementarity: guanine and cytosine; adenine and thymine; and adenine and uracil. As used herein, "complementary" means that the aforementioned relationship applies to substantially all base pairs comprising two single-stranded nucleic acid molecules over the entire length of said molecules. "Partially complementary" refers to the aforementioned relationship in which one of two single-stranded nucleic acid molecules is shorter in length than the other such that a portion of one of the molecules remains single-stranded.

The term "conservative substitution" or "conservative amino acid substitution" refers to a replacement of one or more amino acid residue(s) in a parent protein as stipulated by Table 1.

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"Fragment thereof" refers to a fragment, piece, or sub-region of a nucleic acid or protein molecule whose sequence is disclosed herein, such that said fragment comprises 5 or more amino acids, or 10 or more nucleotides that are contiguous in the parent protein or nucleic acid molecule.

"Functional fragment" as used herein, refers to an isolated sub-region, or fragment of a protein disclosed herein, or sequence of amino acids that, for example,

comprises a functionally distinct region such as an active site for an enzyme, or a binding site for a substrate, or a binding site for a receptor. Functional fragments may be produced by cloning technology, or as the natural products of alternative splicing mechanisms.

"Functionally related" as used herein is applied to proteins or peptides that are predicted to be functionally similar or identical to a progenitor molecule, for example, VEGF-R or fragment thereof. Functionally related species are identified based on chemical and physical similarities in amino acid composition and sequence.

"Host cell" refers to any eucaryotic or procaryotic cell that is suitable for propagating and/or expressing a cloned gene contained on a vector that is introduced into said host cell by, for example, transformation or transfection, or the like.

VEGF-R refers to a gene or cDNA (SEQ ID NO:1) and a protein (SEQ ID NO:2). VEGF-R is a member of the family of VEGF related ligands. This family of molecules is primarily

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responsible for angiogenesis during embryogenesis and in adult tissues, including tumor tissues.

The term "homolog" or "homologous" describes the relationship between different nucleic acid molecules or amino acid sequences in which said sequences or molecules are related by partial identity or similarity at one or more blocks or regions within said molecules or sequences.

The term "hybridization" as used herein refers to a process in which a single-stranded nucleic acid molecule joins with a complementary strand through nucleotide base pairing. "Selective hybridization" refers to hybridization under conditions of high stringency. The degree of hybridization depends upon, for example, the degree of homology, the stringency of hybridization, and the length of hybridizing strands.

"Isolated nucleic acid compound" refers to any RNA or DNA sequence, however constructed or synthesized, which is locationally distinct from its natural location.

A "nucleic acid probe" or "probe" as used herein
is a labeled nucleic acid compound which hybridizes with
another nucleic acid compound. "Nucleic acid probe" means a
single stranded nucleic acid sequence that will combine with
a complementary or partially complementary single stranded
target nucleic acid sequence to form a double-stranded
molecule. A nucleic acid probe may be an oligonucleotide or
a nucleotide polymer. A probe will usually contain a
detectable moiety which may be attached to the end(s) of the
probe or be internal to the sequence of the probe.

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The term "orthologue" or "orthologous" refers to two or more genes or proteins from different organisms that exhibit sequence homology.

The term "paralogue" or "paralogous" refers to two
or more genes or proteins within a single organism that
exhibit sequence homology.

The term "plasmid" refers to an extrachromosomal genetic element. The plasmids disclosed herein are commercially available, publicly available on an unrestricted basis, or can be constructed from readily available plasmids in accordance with published procedures.

A "primer" is a nucleic acid fragment which functions as an initiating substrate for enzymatic or synthetic elongation of, for example, a nucleic acid molecule.

The term "promoter" refers to a nucleic acid sequence that directs transcription, for example, of DNA to RNA. An inducible promoter is one that is regulatable by environmental signals, such as carbon source, heat, or metal ions, for example. A constitutive promoter generally operates at a constant level and is not regulatable.

"Recombinant DNA cloning vector" as used herein refers to any autonomously replicating agent, including, but not limited to, plasmids and phages, comprising a DNA molecule to which one or more additional DNA segments can or have been incorporated.

The term "recombinant DNA expression vector" or
"expression vector" as used herein refers to any recombinant
DNA cloning vector, for example a plasmid or phage, in which
a promoter and other regulatory elements are present thereby

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enabling transcription of an inserted DNA, which may encode a protein.

The term "stringency" refers to hybridization conditions. High stringency conditions disfavor non-homologous base pairing. Low stringency conditions have the opposite effect. Stringency may be altered, for example, by temperature and salt concentration.

"Low stringency" conditions comprise, for example, a temperature of about 37° C or less, a formamide concentration of less than about 50%, and a moderate to low salt (SSC) concentration; or, alternatively, a temperature of about 50° C or less, and a moderate to high salt (SSPE) concentration, for example 1M NaCl.

"High stringency" conditions comprise, for
example, a temperature of about 42° C or less, a formamide concentration of less than about 20%, and a low salt (SSC) concentration; or, alternatively, a temperature of about 65° C, or less, and a low salt (SSPE) concentration. For example, high stringency conditions comprise hybridization in 0.5 M NaHPO4, 7% sodium dodecyl sulfate (SDS), 1 mM EDTA at 65°C (Ausubel, F.M. et al. Current Protocols in Molecular Biology, Vol. I, 1989; Green Inc. New York, at 2.10.3).

"SSC" comprises a hybridization and wash solution.

A stock 20% SSC solution contains 3M sodium chloride, 0.3M sodium citrate, pH 7.0.

"SSPE" comprises a hybridization and wash solution. A 1X SSPE solution contains 180 mM NaCl, 9mM Na₂HPO₄, 0.9 mM NaH₂PO₄ and 1 mM EDTA, pH 7.4.

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"Substantially pure," used in reference to a peptide or protein, means separation from other cellular and non-cellular molecules, including other protein molecules. A substantially pure preparation would be about at least 85% pure; preferably about at least 95% pure. A "substantially pure" protein can be prepared by a variety of techniques, well known to the skilled artisan, including, for example, the IMAC protein purification method.

"Treating" as used herein describes the management
and care of a patient for the purpose of combating the
disease, condition, or disorder and includes the
administration of a protein of the present invention to
prevent the onset of the symptoms or complications,
alleviating the symptoms or complications, or eliminating
the disease, condition, or disorder. Treating as used herein
includes the administration of the protein for cosmetic
purposes. A cosmetic purpose seeks to control, for example,
the weight of a mammal to improve bodily appearance.

The term "vector" as used herein refers to a

20 nucleic acid compound used for introducing exogenous or
endogenous DNA into host cells. A vector comprises a
nucleotide sequence which may encode one or more protein
molecules. Plasmids, cosmids, viruses, and bacteriophages,
in the natural state or which have undergone recombinant
25 engineering, are examples of commonly used vectors.

The various restriction enzymes disclosed and described herein are commercially available and the manner of use of said enzymes including reaction conditions, cofactors, and other requirements for activity are well known to one of ordinary skill in the art. Reaction conditions for

particular enzymes were carried out according to the manufacturer's recommendation.

The VEGF-R gene encodes a novel protein that is related to the VEGF family of proteins. The VEGF-R cDNA comprises a DNA sequence specified herein by SEQ ID NO:1, the coding region being defined by residues 276 through 1310 of SEQ ID NO:1. Those skilled in the art will recognize that owing to the degeneracy of the genetic code, numerous "silent" substitutions of nucleotide base pairs could be introduced into the sequence identified as SEQ ID NO:1 without altering the identity of the encoded amino acid(s) or protein product. All such substitutions are intended to be within the scope of the invention.

Also contemplated by the present invention are VEGF-R proteins, functional fragments thereof, analogs, and 15 functionally related molecules. The VEGF-R protein may be membrane-bound in vivo. For example, expression of a VEGF-R DNA expression vector in 293 T cells or CHO cells showed VEGF-R on the cell membrane of transfected cells as detected by FACS staining using an anti-VEGF-R serum. A secreted form of VEGF-R can be produced by removing a portion of the sequence at the amino terminus and fusing therefore, any suitable signal peptide to facilitate secretion from an expression host cell. For example, residues from about 1 through about 22 of SEQ ID NO:2, or residues from about 1 through 15 of SEQ ID NO:2 can be removed and replaced with the kappa light chain signal sequence, or any other suitable signal peptide, for example the protrypsin signal peptide. Functional fragments comprising sub-regions of VEGF-R (SEQ ID NO:2) are also contemplated by the present invention. One 30

functional fragment comprises from about residues 1 through about 29 of SEQ ID NO:2; another functional fragment comprises from about residues 30 through about 80 of SEQ ID NO:2. These subregions define a kinase domain receptor (KDR) binding site.

Other functional fragments are conveniently identified as—fragments—of—an—intact—VEGF—R—protein_that_retain_the____capacity to induce angiogenesis in vivo or in vitro.

Amino acid substitution modifications can be made in
accordance with the following Table. Modifications of VEGF-R
peptides made in accordance with the Table are generally
expected to retain the biological activity of the parent
molecule based on the art recognized substitutability of
certain amino acids (See e.g. M. Dayhoff, In Atlas of
Protein Sequence and Structure, Vol. 5, Supp. 3, pgs 345352, 1978). Functionality is easily tested in an assay that
measures endothelial cell mitogenic activity, for example.

	T
ORIGINAL RESIDUE	EXEMPLARY SUBSTITUTIONS
ALA	SER, THR
ARG	LYS
ASN	HIS, SER
ASP	GLU, ASN
CYS	SER
GLN	ASN, HIS
GLU	ASP, GLU
GLY	ALA, SER
ніѕ	ASN, GLN
ILE	LEU, VAL, THR
LEU	ILE, VAL
LYS	ARG, GLN, GLU, THR
MET	LEU, ILE, VAL
PHE	LEU, TYR
SER	THR, ALA, ASN
THR	SER, ALA
TRP	ARG, SER
TYR	PHE
VAL	ILE, LEU, ALA
PRO	ALA

Functionally related proteins and peptides

Analogs having biological activities that are similar or identical to VEGF-R, for example, the ability to induce angiogenesis, in vivo or in vitro, are also contemplated by the present invention. Said analogs, while being functionally related, comprise amino acid sequences that

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differ from SEQ ID NO:2. Functional analogs of VEGF-R can be generated by deletion, insertion, inversion, and/or substitution of one or more amino acid residues in said VEGF-R. Substitution analogs can generally be made by solid phase or recombinant techniques in which single or multiple conservative amino acid substitutions are made, for example, according—to—Table—1. Generally,—in—the—case—of—multiple——substitutions, it is preferred that less than ten residues be changed in any given molecule, most preferably between one to five residues are changed in any given molecule such that about between 90% to 99% of residues are identical with the sequence of SEQ ID NO:2; alternatively, such that about between 95% to 99% of residues are identical with SEQ ID NO:2.

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Fragments of proteins

One embodiment of the instant invention provides fragments of the proteins disclosed that may or may not be biologically active. Such fragments are useful, for example, as an antigen for producing an antibody to said proteins.

Fragments of the proteins disclosed herein may be generated by any number of suitable techniques, including chemical synthesis of any portion of SEQ ID NO:2, proteolytic digestion of SEQ ID NO:2, or most preferably, by recombinant DNA mutagenesis techniques, well known to the skilled artisan. See. e.g. K. Struhl, "Reverse biochemistry: Methods and applications for synthesizing yeast proteins in vitro," Meth. Enzymol. 194, 520-535. For example, in a preferred method, a nested set of deletion mutations are introduced into a nucleic acid sequence encoding VEGF-R

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(e.g. residues 276 through 1310 of SEQ ID NO:1) such that varying amounts of the protein coding region are deleted, either from the amino terminal end, or from the carboxyl end of the protein molecule. This method can also be used to create internal fragments of the intact protein in which both the carboxyl and amino terminal ends are removed. Several appropriate nucleases can be used to create such deletions, for example Bal31, or in the case of a single stranded nucleic acid molecule, mung bean nuclease. For simplicity, it is preferred that the VEGF-R gene be cloned into a single-stranded cloning vector, such as bacteriophage M13, or equivalent. If desired, the resulting gene deletion fragments can be subcloned into any suitable vector for propagation and expression of said fragments in any suitable host cell.

Functional fragments of the proteins disclosed herein may be produced as described above, preferably using cloning techniques to engineer smaller versions of the intact gene, lacking sequence from the 5' end, the 3' end, from both ends, or from an internal site. Fragments may be tested for biological activity using any suitable assay, for example, the ability of a protein fragment to induce apoptosis, in vivo or in vitro.

25 Gene Isolation Procedures

Those skilled in the art will recognize that the VEGF-R gene could be obtained by a plurality of recombinant DNA techniques including, for example, hybridization, polymerase chain reaction (PCR) amplification, or de novo DNA

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synthesis. (See e.g., T. Maniatis et al. Molecular Cloning: A Laboratory Manual, 2d Ed. Chap. 14 (1989)).

Methods for constructing cDNA libraries in a suitable vector such as a plasmid or phage for propagation in procaryotic or eucaryotic cells are well known to those skilled in the art. [See e.g. Maniatis et al. Supra].

Suitable cloning vectors—are—well—known—and—are_widely_available.

The VEGF-R gene, or fragment thereof, can be isolated from a tissue in which said gene is expressed, for example, 10 placenta. In one method, mRNA is isolated, and first strand cDNA synthesis is carried out. A second round of DNA synthesis can be carried out for the production of the second strand. If desired, the double-stranded cDNA can be cloned into any suitable vector, for example, a plasmid, 15 thereby forming a cDNA library. Oligonucleotide primers targeted to any suitable region of SEQ ID NO:1 can be used for PCR amplification of VEGF-R. See e.g. PCR Protocols: A Guide to Method and Application, Ed. M. Innis et al., Academic Press (1990). The PCR amplification comprises 20 template DNA, suitable enzymes, primers, and buffers, and is conveniently carried out in a DNA Thermal Cycler (Perkin Elmer Cetus, Norwalk, CT). A positive result is determined by detecting an appropriately-sized DNA fragment following 25 agarose gel electrophoresis.

Protein Production Methods

One embodiment of the present invention relates to the substantially purified protein encoded by the VEGF-R 30 gene.

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Skilled artisans will recognize that the proteins of the present invention can be synthesized by a number of different methods, such as chemical methods well known in the art, including solid phase peptide synthesis or recombinant methods. Both methods are described in U.S. Patent 4,617,149, incorporated herein by reference.

The principles of solid phase chemical synthesis of polypeptides are well known in the art and may be found in general texts in the area. See, e.g., H. Dugas and C. Penney, Bioorganic Chemistry (1981) Springer-Verlag, New York, 54-92. For example, peptides may be synthesized by solid-phase methodology utilizing an Applied Biosystems 430A peptide synthesizer (Applied Biosystems, Foster City, CA) and synthesis cycles supplied by Applied Biosystems.

The proteins of the present invention can also be produced by recombinant DNA methods using the cloned VEGF-R gene. Recombinant methods are preferred if a high yield is desired. Expression of the cloned gene can be carried out in a variety of suitable host cells, well known to those skilled in the art. For this purpose, the VEGF-R gene is introduced into a host cell by any suitable means, well known to those skilled in the art. While chromosomal integration of the cloned gene is within the scope of the present invention, it is preferred that the gene be cloned into a suitable extra-chromosomally maintained expression vector so that the coding region of the VEGF-R gene is operably-linked to a constitutive or inducible promoter.

The basic steps in the recombinant production of the VEGF-R protein are:

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- a) constructing a natural, synthetic or semi-synthetic DNA encoding VEGF-R protein;
- b) integrating said DNA into an expression vector in a manner suitable for expressing the VEGF-R protein, either alone or as a fusion-protein;
 - c) transforming or otherwise introducing said vector into an appropriate eucaryotic or prokaryotic host cell forming a recombinant host cell,
 - d) culturing said recombinant host cell in a manner to express the VEGF-R protein; and
 - e) recovering and substantially purifying the VEGF-R protein by any suitable means, well known to those skilled in the art.

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Expressing Recombinant VEGF-R Protein in Procaryotic and Eucaryotic Host Cells

Procaryotes may be employed in the production of recombinant VEGF-R protein. For example, the *Escherichia coli* K12 strain 294 (ATCC No. 31446) is particularly useful for the prokaryotic expression of foreign proteins. Other strains of *E. coli*, bacilli such as *Bacillus subtilis*, enterobacteriaceae such as *Salmonella typhimurium* or *Serratia marcescans*, various Pseudomonas species and other bacteria, such as *Streptomyces*, may also be employed as host

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cells in the cloning and expression of the recombinant proteins of this invention.

Promoter sequences suitable for driving the expression of genes in procaryotes include b -lactamase [e.g. vector pGX2907, ATCC 39344, contains a replicon and b -lactamase gene], lactose systems [Chang et al., Nature (London), 275:615 (1978); Goeddel et al., Nature (London), 281:544 (1979)], alkaline phosphatase, and the tryptophan (trp) promoter system [vector pATH1 (ATCC 37695)], which is designed to facilitate expression of an open reading frame 10 as a trpE fusion protein under the control of the trp promoter. Hybrid promoters such as the tac promoter (isolatable from plasmid pDR540, ATCC-37282) are also Still other bacterial promoters, whose nucleotide suitable. 15 sequences are generally known, may be ligated to DNA encoding the protein of the instant invention, using linkers or adapters to supply any required restriction sites. Promoters for use in bacterial systems also will contain a Shine-Dalgarno sequence operably-linked to the DNA encoding 20 the desired polypeptides. These examples are illustrative rather than limiting.

either by direct expression or as a fusion protein comprising the protein of interest as a translational fusion with another protein or peptide which may be removable by enzymatic or chemical cleavage. It is often observed in the production of certain peptides in recombinant systems that expression as a fusion protein prolongs the life span, increases the yield of the desired peptide, or provides a convenient means of purifying the protein. This is

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particularly relevant when expressing mammalian proteins in procaryotic hosts. A variety of peptidases (e.g. enterokinase and thrombin) which cleave a polypeptide at specific sites or digest the peptides from the amino or carboxy termini (e.g. diaminopeptidase) of the peptide chain are known. Furthermore, particular chemicals (e.g. cyanogen bromide)—will-cleave—a—polypeptide—chain—at—specific_sites.—The skilled artisan will appreciate the modifications necessary to the amino acid sequence (and synthetic or semisynthetic coding sequence if recombinant means are employed) to incorporate site-specific internal cleavage sites. See e.g., P. Carter, "Site Specific Proteolysis of Fusion Proteins", Chapter 13, in Protein Purification: From Molecular Mechanisms to Large Scale Processes, American Chemical Society, Washington, D.C. (1990).

In addition to procaryotes, a variety of amphibian expression systems such as frog oocytes, and mammalian cell systems can be used. The choice of a particular host cell depends to some extent on the particular expression vector used. Exemplary mammalian host cells suitable for use in the present invention include HepG-2 (ATCC HB 8065), CV-1 (ATCC CCL 70), LC-MK2 (ATCC CCL 7.1), 3T3 (ATCC CCL 92), CHO-K1 (ATCC CCL 61), HeLa (ATCC CCL 2), RPMI8226 (ATCC CCL 155), H4IIEC3 (ATCC CCL 1600), C127I (ATCC CCL 1616), HS-Sultan (ATCC CCL 1484), and BHK-21 (ATCC CCL 10), for example.

A wide variety of vectors are suitable for transforming mammalian host cells. For example, the pSV2-type vectors comprise segments of the simian virus 40 (SV40) genome required for transcription and polyadenylation. A

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large number of plasmid pSV2-type vectors have been constructed, such as pSV2-gpt, pSV2-neo, pSV2-dhfr, pSV2-hyg, and pSV2-b-globin, in which the SV40 promoter drives transcription of an inserted gene. These vectors are widely available from sources such as the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland, 20852, or the Northern Regional Research Laboratory (NRRL), 1815 N. University Street, Peoria, Illinois, 61604.

Promoters suitable for expression in mammalian cells include the SV40 late promoter, promoters from eukaryotic genes, such as, for example, the estrogen-inducible chicken ovalbumin gene, the interferon genes, the glucocorticoid-inducible tyrosine aminotransferase gene, the thymidine kinase gene promoter, and the promoters of the major early and late adenovirus genes and the cytomegalovirus promoter.

Plasmid pRSVcat (ATCC 37152) comprises portions of a long terminal repeat of the Rous Sarcoma virus, a virus known to infect chickens and other host cells. This long 20 terminal repeat contains a promoter which is suitable for use in the vectors of this invention. H. Gorman et al., Proc. Nat. Acad. Sci. (USA), 79, 6777 (1982). The plasmid pMSVi (NRRL B-15929) comprises the long terminal repeats of 25 the Murine Sarcoma virus, a virus known to infect mouse and other host cells. The mouse metallothionein promoter has also been well characterized for use in eukaryotic host cells and is suitable for use in the present invention. This promoter is present in the plasmid pdBPV-MMTneo (ATCC

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37224) which can serve as the starting material for the construction of other plasmids of the present invention.

Transfection of mammalian cells with vectors can be performed by a plurality of well known processes including, but not limited to, protoplast fusion, calcium phosphate co-precipitation, electroporation and the like.

See, e.g., Maniatis et al., supra.

Some viruses also make appropriate vectors.

Examples include the adenoviruses, the adeno-associated viruses, the vaccinia virus, the herpes viruses, the baculoviruses, and the rous sarcoma virus, as described in U.S. Patent 4,775,624, incorporated herein by reference.

Eucaryotic microorganisms such as yeast and other fungi are also suitable host cells. The yeast Saccharomyces cerevisiae is the preferred eucaryotic microorganism. Other yeasts such as Kluyveromyces lactis and Pichia pastoris are also suitable. For expression in Saccharomyces, the plasmid YRp7 (ATCC-40053), for example, may be used. See, e.g., L. Stinchcomb et al., Nature, 282, 39 (1979); J. Kingsman et al., Gene, 7, 141 (1979); S. Tschemper et al., Gene, 10, 157 (1980). Plasmid YRp7 contains the TRP1 gene which provides a selectable marker for use in a trp1 auxotrophic mutant.

Purification of Recombinantly-Produced VEGF-R Protein

25 An expression vector carrying the cloned VEGF-R gene is transformed or transfected into a suitable host cell using standard methods. Cells that contain the vector are propagated under conditions suitable for expression of the recombinant VEGF-R protein. For Example, if the recombinant 30 gene has been placed under the control of an inducible

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promoter, suitable growth conditions would incorporate the appropriate inducer. The recombinantly-produced protein may be purified from cellular extracts of transformed cells by any suitable means.

In a preferred process for protein purification, the VEGF-R gene is modified at the 5' end to incorporate several histidine residues at the amino terminus of the VEGF-R protein. This "histidine tag" enables a single-step protein purification method referred to as "immobilized metal ion affinity chromatography" (IMAC), essentially as described in U.S. Patent 4,569,794, which hereby is incorporated by reference. The IMAC method enables rapid isolation of substantially pure recombinant VEGF-R protein starting from a crude extract of cells that express a modified recombinant protein, as described above.

Production of Antibodies

The proteins of this invention and fragments
thereof may be used in the production of antibodies. The
term "antibody" as used herein describes antibodies,

fragments of antibodies (such as, but not limited, to Fab,
Fab', Fab2', and Fv fragments), and chimeric, humanized,
veneered, resurfaced, or CDR-grafted antibodies capable of
binding antigens of a similar nature as the parent antibody
molecule from which they are derived. The instant invention
also encompasses single chain polypeptide binding molecules.

The production of antibodies, both monoclonal and polyclonal, in animals, especially mice, is well known in the art. See, e.g., C. Milstein, Handbook of Experimental Immunology, (Blackwell Scientific Pub., 1986); J. Goding, Monoclonal Antibodies: Principles and Practice, (Academic

Press, 1983). For the production of monoclonal antibodies the basic process begins with injecting a mouse, or other suitable animal, with an immunogen. The mouse is subsequently sacrificed and cells taken from its spleen are fused with myeloma cells, resulting in a hybridoma that reproduces in vitro. The population of hybridomas is screened to isolate individual clones, each of which secretes a single antibody species, specific for the immunogen. Each antibody obtained in this way is the clonal product of a single B cell.

Chimeric antibodies are described in U.S. Patent No. 4,816,567, the entire contents of which is herein incorporated by reference. This reference discloses methods and vectors for the preparation of chimeric antibodies. An alternative approach is provided in U.S. Patent No. 4,816,397, the entire contents of which is herein incorporated by reference. This patent teaches coexpression of the heavy and light chains of an antibody in the same host cell.

The approach of U.S. Patent 4,816,397 has been further refined in European Patent Publication No. 0 239 400. The teachings of this European patent publication are a preferred format for genetic engineering of monoclonal antibodies. In this technology the complementarity determining regions (CDRs) of a human antibody are replaced with the CDRs of a murine monoclonal antibody, thereby converting the specificity of the human antibody to the specificity of a murine antibody.

Single chain antibodies and libraries thereof are
yet another variety of genetically engineered antibody

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technology that is well known in the art. (See, e.g. R.E. Bird, et al., Science 242:423-426 (1988); PCT Publication Nos. WO 88/01649, WO 90/14430, and WO 91/10737. Single chain antibody technology involves covalently joining the binding regions of heavy and light chains to generate a single polypeptide chain. The binding specificity of the intact antibody molecule is thereby reproduced on a single polypeptide chain.

The antibodies contemplated by this invention are useful in diagnostics, therapeutics or in diagnostic/therapeutic combinations.

The proteins of this invention or suitable fragments thereof can be used to generate polyclonal or monoclonal antibodies, and various inter-species hybrids, or humanized antibodies, or antibody fragments, or single-chain antibodies. The techniques for producing antibodies are well known to skilled artisans. (See e.g. A.M. Campbell, Monoclonal Antibody Technology: Laboratory Techniques in Biochemsitry and Molecular Biology, Elsevier Science Publishers, Amsterdam (1984); Kohler and Milstein, Nature 256, 495-497 (1975); Monoclonal Antibodies: Principles & Applications Ed. J.R.Birch & E.S. Lennox, Wiley-Liss, 1995.

A protein used as an immunogen may be modified or administered in an adjuvant, by subcutaneous or

intraperitoneal injection into, for example, a mouse or a rabbit. For the production of monoclonal antibodies, spleen cells from immunized animals are removed, fused with myeloma cells, such as SP2/0-Ag14 cells, and allowed to become monoclonal antibody producing hybridoma cells in the manner known to the skilled artisan. Hybridomas that secrete a

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desired antibody molecule can be screened by a variety of well known methods, for example ELISA assay, western blot analysis, or radioimmunoassay (Lutz et al. Exp. Cell Res. 175, 109-124 (1988); Monoclonal Antibodies: Principles & Applications Ed. J.R.Birch & E.S. Lennox, Wiley-Liss, 1995).

For some applications labeled antibodies are desirable.

Procedures for labeling-antibody-molecules-are-widely-known, including for example, the use of radioisotopes, affinity labels, such as biotin or avidin, enzymatic labels, for example horseradish peroxidase, and fluorescent labels, such as FITC or rhodamine (See e.g. Enzyme-Mediated Immunoassay, Ed. T. Ngo, H. Lenhoff, Plenum Press 1985; Principles of Immunology and Immunodiagnostics, R.M. Aloisi, Lea & Febiger, 1988).

Labeled antibodies are useful for a variety of diagnostic applications. In one embodiment the present invention relates to the use of labeled antibodies to detect the presence of VEGF-R. Alternatively, the antibodies could be used in a screen to identify potential modulators of VEGF-R. For example, in a competitive displacement assay, the antibody or compound to be tested is labeled by any suitable method. Competitive displacement of an antibody from an antibody-antigen complex by a test compound such that a test compound-antigen complex is formed provides a method for identifying compounds that bind VEGF-R.

Other embodiments of the present invention comprise isolated nucleic acid sequences that encode SEQ ID NO:2, or related nucleic acids that are at least about 75% identical to SEQ ID NO:1, or to their complementary sequence, or nucleic acids that hybridize to SEQ ID NO:1

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under high stringency conditions. Such sequences may come, for example, from paralogous or orthologous genes.

The VEGF-R cDNA (viz. SEQ ID NO:1) and related nucleic acid molecules that encode SEQ ID NO:2, or functional fragments thereof, may be produced by chemical synthetic methods. The synthesis of nucleic acids is well known in the art. See, e.g., E.L. Brown, R. Belagaje, M.J. Ryan, and H.G. Khorana, Methods in Enzymology, 68:109-151 (1979). Fragments of the DNA sequence corresponding to the VEGF-R gene could be generated using a conventional DNA 10 synthesizing apparatus, such as the Applied Biosystems Model 380A or 380B DNA synthesizers (Applied Biosystems, Inc., 850 Lincoln Center Drive, Foster City, CA 94404) using phosphoramidite chemistry, thereafter ligating the fragments so as to reconstitute the entire gene. Alternatively, 15 phosphotriester chemistry may be employed to synthesize the nucleic acids of this invention. (See, e.g., M.J. Gait, ed., Oligonucleotide Synthesis, A Practical Approach, (1984)).

In an alternative methodology, namely PCR, the
DNA sequences disclosed and described herein, comprising,
for example, a portion or all of SEQ ID NO:1 can be produced
from a plurality of starting materials. For example,
starting with a cDNA preparation (e.g. cDNA library) derived
from a tissue that expresses the VEGF-R gene, suitable
oligonucleotide primers complementary to SEQ ID NO:1 or to
any sub-region therein, are prepared as described in U.S.
Patent No. 4,889,818, hereby incorporated by reference.
Other suitable protocols for the PCR are disclosed in PCR
Protocols: A Guide to Method and Applications, Ed. Michael

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A. Innis et al., Academic Press, Inc. (1990). Using PCR, any region of the VEGF-R gene can be targeted for amplification such that full or partial length gene sequences may be produced.

The ribonucleic acids of the present invention may be prepared using polynucleotide synthetic methods discussed supra, or they may be prepared enzymatically, for example, using RNA polymerase to transcribe a VEGF-R DNA template.

The most preferred systems for preparing the ribonucleic acids of the present invention employ the RNA polymerase from the bacteriophage T7 or the bacteriophage SP6. These RNA polymerases are highly specific, requiring the insertion of bacteriophage-specific sequences at the 5' end of the template to be transcribed. See, Maniatis et al., supra.

This invention also provides nucleic acids, RNA or DNA, that are complementary to SEQ ID NO:1, or fragment thereof.

Nucleic Acid Probes

The present invention also provides probes and primers useful for a variety of molecular biology techniques including, for example, hybridization screens of genomic, subgenomic, or cDNA libraries, as well as hybridization against nucleic acids derived from cell lines or tissues that originate from drug-resistant tumors. Such hybridization screens are useful as methods to identify homologous and/or functionally related sequences from the same or other organisms. A nucleic acid compound comprising SEQ ID NO:1, or a complementary sequence thereof, or a fragment thereof, which is at least 14 base pairs in length,

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and which will selectively hybridize to human DNA or mRNA encoding VEGF-R protein or fragment thereof, or a functionally related protein, is provided. Preferably, the 14 or more base pair compound is DNA. See e.g. B. Wallace and G. Miyada, "Oligonucleotide Probes for the Screening of Recombinant DNA Libraries," In Meth. Enzym., 152, 432-442, Academic Press (1987).

Probes and primers can be prepared by enzymatic or recombinant methods, well known to those skilled in the art (See e.g. Sambrook et al. supra). A probe may be a single stranded nucleic acid sequence which is complementary in some particular degree to a nucleic acid sequence sought to be detected ("target sequence"). A probe may be labeled with a detectable moiety such as a radio-isotope, antigen, or chemiluminescent moiety. A description of the use of nucleic acid hybridization as a procedure for the detection of particular nucleic acid sequences is described in U.S. Patent No. 4,851,330 to Kohne, entitled "Method for Detection, Identification and Quantitation of Non-Viral Organisms."

Having the DNA sequence of the present invention allows preparation of relatively short DNA (or RNA) sequences having the ability to specifically hybridize to gene sequences disclosed herein. In these aspects, nucleic acid probes of an appropriate length are prepared. The ability of such nucleic acid probes to specifically hybridize to a polynucleotide encoding a VEGF-R gene or related sequence lends particular utility in a variety of embodiments. Most importantly, the probes may be used in a variety of assays

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for detecting the presence of complementary sequences in a given sample.

In certain embodiments, it is advantageous to use oligonucleotide primers. The sequence of such primers is designed using a polynucleotide of the present invention for use in detecting, amplifying or mutating a defined segment of a gene or polynucleotide that encodes a VEGF-R polypeptide using PCR technology.

Preferred nucleic acid sequences employed for hybridization studies, or assays, include probe molecules 10 that are complementary to at least an about 14 to an about 70-nucleotide long stretch of a polynucleotide that encodes a VEGF-R polypeptide, such as the nucleotide base sequences designated as SEQ ID NO:1. A 15 length of at least 14 nucleotides helps to ensure that the fragment is of sufficient length to form a duplex molecule that is both stable and selective. Molecules having complementary sequences over stretches greater than 14 bases in length are generally preferred, though in order to 20 increase stability and selectivity of the hybrid. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 25 to 40 nucleotides, 55 to 70 nucleotides, or even longer where desired. Such fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, by application 25 of nucleic acid reproduction technology, such as the PCR TM technology of U.S. Pat. No. 4,603,102, herein incorporated by reference, or by excising selected DNA fragments from recombinant plasmids containing appropriate inserts and 30 suitable restriction enzyme sites.

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The following guidelines are useful for designing probes with desirable characteristics. The extent and specificity of hybridization reactions are affected by a number of factors that determine the sensitivity and specificity of a particular probe, whether perfectly complementary to its target or not. The affect of various experimental parameters and conditions are well known to those skilled in the art.

First, the stability of the probe:target nucleic acid hybrid should be chosen to be compatible with the assay conditions. This may be accomplished by avoiding long A and T rich sequerces, by terminating the hybrids with G:C base pairs and by designing a probe with an appropriate Tm (i.e. melting temperature). The melting profile, including the Tm of a hybrid comprising an oligonucleotide and target sequence, may be determined using a Hybridization Protection Assay. The probe should be chosen so that the length and % GC content result in a Tm about 2°-10° C higher than the temperature at which the final assay will be performed. The base composition of the probe is also a significant factor because G-C base pairs exhibit greater thermal stability as compared to A-T base pairs. Thus, hybridization involving complementary nucleic acids of higher G-C content will be more stable at higher temperatures.

The ionic strength and incubation temperature under which a probe will be used should also be taken into account. It is known that hybridization will increase as the ionic strength of the reaction mixture increases, and that the thermal stability of molecular hybrids will increase with increasing ionic strength. On the other hand, chemical

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reagents such as formamide, urea, DMSO and alcohols, which disrupt hydrogen bonds, increase the stringency of hybridization. Destabilization of hydrogen bonds by such reagents can greatly reduce the Tm. In general, optimal hybridization for synthetic oligonucleotide probes of about 10-50 bases in length occurs approximately 5° C below the melting temperature for a given duplex. Incubation at temperatures below the optimum may allow mismatched base sequences to hybridize and can therefore result in reduced specificity.

The length of the target nucleic acid sequence and, accordingly, the length of the probe sequence can also be important. In some cases, there may be several sequences from a particular region, varying in location and length, which will yield probes with the desired hybridization characteristics. In other cases, one sequence may be significantly better than another even though the one sequence differs merely by a single base. Finally, there can be intramolecular and intermolecular hybrids formed within a probe if there is sufficient self-complementarity. Such structures can be avoided through careful probe design. Computer programs are available to search for this type of interaction.

A probe molecule may be used for hybridizing to a

25 sample suspected of possessing a VEGF-R or VEGF-R-related
nucleotide sequence. The hybridization reaction is carried
out under suitable conditions of stringency.

Alternatively, such DNA molecules may be used in a number of techniques including their use as: (1) diagnostic tools to detect polymorphisms in DNA samples from a human or

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other mammal; (2) means for detecting and isolating homologs of VEGF-R and related polypeptides from a DNA library potentially containing such sequences; (3) primers for hybridizing to related sequences for the purpose of amplifying those sequences; and (4) primers for altering the native VEGF-R DNA sequences; as well as other techniques which rely on the similarity of the DNA sequences to those of the VEGF-R DNA segments herein disclosed.

Once synthesized, oligonucleotide probes may be labeled 10 by any of several well known methods. See e.q. Maniatis et.al., Molecular Cloning (2d ed. 1989). Useful labels include radioisotopes, as well as non-radioactive reporting groups. Isotopic labels include H3, S35, P32, I125, Cobalt, and C14. Most methods of isotopic labeling involve the use of enzymes and include methods such as nick-translation, end-15 labeling, second strand synthesis, and reverse transcription. When using radio-labeled probes, hybridization can be detected by autoradiography, scintillation counting, or gamma counting. The detection 20 method selected will depend upon the hybridization conditions and the particular radio isotope used for labeling.

Non-isotopic materials can also be used for labeling, and may be introduced internally into the sequence or at the end of the sequence. Modified nucleotides may be incorporated enzymatically or chemically, and chemical modifications of the probe may be performed during or after synthesis of the probe, for example, by the use of non-nucleotide linker groups. Non-isotopic labels include

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fluorescent molecules, chemiluminescent molecules, enzymes, cofactors, enzyme substrates, haptens or other ligands.

In a preferred embodiment of the invention, the length of an oligonucleotide probe is greater than or equal to about 18 nucleotides and less than or equal to about 50 nucleotides.

Labeling of an oligonucleotide of the present invention may be performed enzymatically using [32P] -labeled ATP and the enzyme T4 polynucleotide kinase.

Vectors

Another aspect of the present invention relates to recombinant DNA cloning vectors and expression vectors comprising the nucleic acids of the present invention. The preferred nucleic acid vectors are those which comprise DNA. The most preferred recombinant DNA vectors comprise the isolated DNA sequence, defined by residues 276 through 1310 of SEQ ID NO:1.

The skilled artisan understands that choosing the most appropriate cloning vector or expression vector depends upon a number of factors including the availability of restriction enzyme sites, the type of host cell into which the vector is to be transfected or transformed, the purpose of the transfection or transformation (e.g., stable transformation as an extrachromosomal element, or integration into the host chromosome), the presence or absence of readily assayable or selectable markers (e.g., antibiotic resistance and metabolic markers of one type and another), and the number of copies of the gene desired in the host cell.

Vectors suitable to carry the nucleic acids of the present invention comprise RNA viruses, DNA viruses, lytic

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bacteriophages, lysogenic bacteriophages, stable bacteriophages, plasmids, viroids, and the like. The most preferred vectors are plasmids.

When preparing an expression vector the skilled 5 artisan understands that there are many variables to be considered, for example, whether to use a constitutive or inducible promoter. The practitioner also understands that the amount of nucleic acid or protein to be produced dictates, in part, the selection of the expression system. Regarding promoter sequences, inducible promoters are 10 preferred because they enable high level, regulatable expression of an operably-linked gene. The skilled artisan will recognize a number of suitable promoters that respond to a variety of inducers, for example, carbon source, metal 15 ions, and heat. Other relevant considerations regarding an expression vector include whether to include sequences for directing the localization of a recombinant protein. For example, a sequence encoding a signal peptide preceding the coding region of a gene is useful for directing the extra-20 cellular export of a resulting polypeptide.

The present invention also provides a method for constructing a recombinant host cell capable of expressing VEGF-R proteins (e.g. SEQ ID NO:2), said method comprising transforming or otherwise introducing into a host cell a recombinant DNA vector that comprises an isolated DNA sequence that encodes SEQ ID NO:2. A suitable host cell is any eucaryotic cell that can accommodate high level expression of an exogenously introduced gene or protein, and that will incorporate said protein into its membrane structure. Vectors for expression are those which comprise

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SEQ ID NO:1 or fragment thereof. Transformed host cells may be cultured under conditions well known to skilled artisans such that SEQ ID NO:2 is expressed, thereby producing a recombinant VEGF-R protein in the recombinant host cell.

For the purpose of identifying compounds having utility as inhibitors of tumor growth, for example, in a treatment of cancer, it would be desirable to identify compounds that bind the VEGF-R protein and/or modify or antagonize its activity. A method for determining agents that bind the VEGF-R protein comprises contacting the VEGF-R protein with a test compound and monitoring binding by any suitable means.

The instant invention provides a screening system for discovering compounds that bind the VEGF-R protein, said screening system comprising the steps of:

- a) preparing VEGF-R protein;
- b) exposing said VEGF-R protein to a test compound;
- c) quantifying the binding of said compound to VEGF-R protein by any suitable means.

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Utilization of the screening system described above provides a means to determine compounds that may alter the biological function of VEGF-R. This screening method may be adapted to large-scale, automated procedures such as a PANDEX® (Baxter-Dade Diagnostics) system, allowing for efficient high-volume screening of potential therapeutic agents.

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In such a screening protocol VEGF-R is prepared as described herein, preferably using recombinant DNA technology. A test compound is introduced into a reaction vessel containing the VEGF-R protein or fragment thereof. Binding of VEGF-R by a test compound is determined by any suitable means. For example, in one method radioactivelylabeled or chemically-labeled test compound may be used. Binding of the protein by the compound is assessed, for example, by quantifying bound label versus unbound label using any suitable method. Binding of a test compound may also be carried out by a method disclosed in U.S. Patent 5,585,277, which hereby is incorporated by reference. In this method, binding of a test compound to a protein is assessed by monitoring the ratio of folded protein to unfolded protein, for example by monitoring sensitivity of said protein to a protease, or amenability to binding of said protein by a specific antibody against the folded state of the protein.

identifying a ligand, for example, an antagonist of a VEGF-R protein, as a lead to a pharmaceutical compound for the treatment of cancer, or for inhibiting tumor growth. A ligand that binds VEGF-R, or related fragment thereof, is identified, for example, by combining a test ligand with

VEGF-R under conditions that cause the protein to exist in a ratio of folded to unfolded states. If the test ligand binds the folded state of the protein, the relative amount of folded protein will be higher than in the case of a test ligand that does not bind the protein. The ratio of protein in the folded versus unfolded state is easily determinable

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by, for example, susceptibility to digestion by a protease, or binding to a specific antibody, or binding to chaperonin protein, or binding to any suitable surface.

VEGF-R Protein Therapeutic Applications

In one embodiment, the present invention relates to therapeutic applications in which angiogenesis or the inhibition thereof is therapeutically beneficial. For example, VEGF-R may be administered to enhance wound healing such as in post-surgical procedures, or in the treatment of cardiac vessel blockage, or other condition in which enhanced blood vessel formation is beneficial.

The present invention also provides methods for treating cancer and for inhibiting tumor growth, in vitro or in vivo, comprising administration of an effective amount of an VEGF-R antagonist.

For therapeutic use in wound healing or other use in which enhanced angiogenesis is advantageous, an effective amount of VEGF-R protein is administered to an organism in need thereof in a dose between about 0.1 and 1000 ug/kg body weight. In practicing the methods contemplated, VEGF-R can be administered in a single daily dose or in multiple doses per day. The amount per administration will be determined by the physician and depend on such factors as the nature and severity of the disease, and the age and general health of the patient.

The present invention also provides a pharmaceutical composition comprising as the active agent a VEGF-R polypeptide or fragment thereof, or a pharmaceutically acceptable non-toxic salt thereof, and a pharmaceutically acceptable solid or liquid carrier. For

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example, compounds comprising VEGF-R can be admixed with conventional pharmaceutical carriers and excipients, and used in the form of tablets, capsules, elixirs, suspensions, syrups, wafers, and the like. The compositions comprising VEGF-R will contain from about 0.1% to 90% by weight of the active compound, and more generally from about 10% to 30%. The compositions may contain common carriers and excipients such as corn starch or gelatin, lactose, sucrose, microcrystalline cellulose, kaolin, mannitol, dicalcium phosphate, sodium chloride, and alginic acid. The compounds can be formulated for oral or parenteral administration.

For intravenous (IV) use, the VEGF-R protein is administered in commonly used intravenous fluid(s) and administered by infusion. Such fluids, for example, physiological saline, Ringer's solution or 5% dextrose solution can be used.

For intramuscular preparations, a sterile formulation, preferably a suitable soluble salt form of the VEGF-R protein, for example SEQ ID NO:2, such as the hydrochloride salt, can be dissolved and administered in a pharmaceutical diluent such as pyrogen-free water (distilled), physiological saline or 5% glucose solution. A suitable insoluble form of the compound may be prepared and administered as a suspension in an aqueous base or a pharmaceutically acceptable oil base, e.g. an ester of a long chain fatty acid such as ethyl oleate.

The present invention also provides a pharmaceutical composition comprising as the active agent an antagonist of VEGF-R, or a pharmaceutically acceptable non-toxic salt thereof, and a pharmaceutically acceptable solid

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or liquid carrier. For example, compounds comprising VEGF-R antagonist can be admixed with conventional pharmaceutical carriers and excipients, and used in the form of tablets, capsules, elixirs, suspensions, syrups, wafers, and the like. The compositions comprising VEGF-R will contain from about 0.1% to 90% by weight of the active compound, and more generally—from—about—10%—to 30%. The compositions may contain common carriers and excipients such as corn starch or gelatin, lactose, sucrose, microcrystalline cellulose, kaolin, mannitol, dicalcium phosphate, sodium chloride, and alginic acid. The compounds can be formulated for oral or parenteral administration.

Skilled artisans will recognize that IC50 values are dependent on the selectivity of the compound tested.

For example, a compound with an IC50 which is less than 10 nM is generally considered an excellent candidate for drug therapy. However, a compound which has a lower affinity, but is selective for a particular target, may be an even better candidate. The skilled artisan will recognize that any information regarding the binding potential, inhibitory activity, or selectivity of a particular compound is useful toward the development of pharmaceutical products.

The following examples more fully describe the present invention. Those skilled in the art will recognize that the particular reagents, equipment, and procedures described are merely illustrative and are not intended to limit the present invention in any manner.

EXAMPLE 1

RT-PCR Amplification of VEGF-R Gene from mRNA

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A VEGF-R gene is isolated by reverse transcriptase PCR (RT-PCR) using conventional methods. Total RNA from a tissue that expresses the VEGF-R gene, is prepared using standard methods. First strand cDNA synthesis is achieved using a commercially available kit (SuperScript™ System; Life Technologies) in conjunction with specific primers directed at any suitable region of SEQ ID NO:1, for example between residues 276 and 1310.

Amplification is carried out by adding to the first strand cDNA (dried under vacuum): 8 µl of 10X synthesis buffer (200 mM Tris-HCl, pH 8.4; 500 mM KCl, 25 mM MgCl₂, 1 ug/ul BSA); 68 µl distilled water; 1 µl each of a 10 uM solution of each primer; and 1 µl Taq DNA polymerase (2 to 5 U/µl). The reaction is heated at 94° C for 5 min. to denature the RNA/cDNA hybrid. Then, 15 to 30 cycles of PCR amplification are performed using any suitable thermal cycle apparatus. The amplified sample may be analyzed by agarose gel electrophoresis to check for an appropriately-sized fragment.

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EXAMPLE 2

Production of a Vector for Expressing VEGF-R in a Host Cell

An expression vector suitable for expressing VEGF-R or fragment thereof in a variety of procaryotic host cells, such as *E. coli* is easily made. The vector contains an origin of replication (Ori), an ampicillin resistance gene (Amp) useful for selecting cells which have incorporated the vector following a tranformation procedure, and further comprises the T7 promoter and T7 terminator sequences in operable linkage to a VEGF-R coding region.

Plasmid pET11A (obtained from Novogen, Madison WI) is a suitable parent plasmid. pET11A is linearized by restriction with endonucleases NdeI and BamHI. Linearized pET11A is ligated to a DNA fragment bearing NdeI and BamHI sticky ends and comprising the coding region of the VEGF-R gene as disclosed by SEQ ID NO:1 viz. Residues 276 through 1310, or fragment thereof.

The VEGF-R gene used in this construction may be slightly modified at the 5' end (amino terminus of encoded protein) in order to simplify purification of the encoded protein product. For this purpose, an oligonucleotide encoding 8 histidine residues is inserted after the ATG start codon. Placement of the histidine residues at the amino terminus of the encoded protein serves to enable the IMAC one-step protein purification procedure.

EXAMPLE 3

Recombinant Expression of Modified VEGF-R Protein

An expression vector that carries an ORF encoding residues 16 though 345 of SEQ ID NO:2 and further comprising a kappa chain signal peptide fused at the amino terminus of said residue 16 was operably-linked to an CMV promoter in plasmid pcDNA3 and transfected into 293 T cells using standard methods. Transfectants were analyzed for transient expression of VEGF-R using Western blot analysis. The results showed that the truncated VEGF-R protein was secreted into the culture medium.

EXAMPLE 4

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Detecting Ligands that Bind VEGF-R Using a Chaperonin Protein Assay

The wells of an ELISA plate are coated with chaperonin by incubation for several hours with a 4 uq/ml solution of the protein in Tris-buffered Saline (TBS: 10 mM Tris-HCl. pH7.5, 0.2M NaCl). The plates are then washed 3 times with TBS containing 0.1% Tween-20 (TBST). Then, a mixture of VEGF-R protein (sufficient amount to saturate about 50% of the binding sites on chaperonin) and test compound (10'9 to 10^{-5} M) in about 50 μl volume is added to each well of the plate for an incubation of about 60 minutes. Aliquots of the well solutions are then transferred to the wells of fresh plates and incubated for 60 minutes at room temperature, followed by 3 washes with TBST. Next, about 50 μ l of an antibody specific for VEGF-R plus 5% nonfat dry milk are added to each well for a 30 minute incubation at room temperature. After washing, about 50 µl of goat anti-rabbit IgG alkaline phosphatase conjugate at an appropriate dilution in TBST plus 5% nonfat dry milk are added to each will and incubated 30 minutes at room temperature. The plates are washed again with TBST and 0.1 ml of 1 mg/ml pnitrophenylphosphate in 0.1% diethanolamine is added. Color development (proportional to bound alkaline phosphatase antibody conjugate) is monitored with an ELISA plate reader. When test ligand binding has occurred, ELISA analysis reveals VEGF-R in solution at higher concentrations than in the absence of test ligand.

30 EXAMPLE 5

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Production of an Antibody to VEGF-R Protein

Substantially pure VEGF-R protein or fragment thereof is isolated from transfected or transformed cells using any of the well known methods in the art, or by a method specifically disclosed herein. Concentration of protein in a final preparation is adjusted, for example, by filtration through—an—Amicon—filter—device—such—that—the—level—is—about—1 to 5 ug/ml. Monoclonal or polyclonal antibody can be prepared as follows.

Monoclonal antibody can be prepared from murine hybridomas according to the method of Kohler and Milstein (Nature, 256, 495, 1975), or a modified method thereof. Briefly, a mouse is repetitively inoculated with a few micrograms of the protein or fragment thereof, or fusion peptide thereof, over a period of a few weeks. The mouse is then sacrificed and the antibody producing cells of the spleen isolated. The spleen cells are fused by means of polyethylene glycol with mouse myeloma cells. Fused cells that produce antibody are identified by any suitable immunoassay, for example, ELISA, as described in E. Engvall, Meth. Enzymol., 70, 419, 1980.

Polyclonal antiserum can be prepared by well known methods (See e.g. J. Vaitukaitis et.al. Clin. Endocirnol. Metab. 33, 988, 1971) that involve immunizing suitable animals with the proteins, fragments thereof, or fusion proteins thereof, disclosed herein. Small doses (e.g. nanogram amounts) of antigen administered at multiple intradermal sites appears to be the most reliable method.

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Endothelial Cell Growth Assay

A recombinant plasmid that expresses VEGF-R, or functionally related fragment, is transfected by calcium phosphate precipitation into human embryonic kidney cell line 293, cultured in Dulbecco's Modified Eagle's Medium (DMEM)F-12 (1:1) containing 2 mM glutamine and 10% fetal bovine serum. After overnight incubation with the calcium phosphate-DNA mixture, the medium is replaced with serum-free medium and incubated for an additional 48-72 hours before assaying the medium for biological activity.

For conducting the cell growth assay, bovine capillary endothelial cells are maintained in DMEM containing 20% calf serum according to the method of Ferrara, Biochem. Biophys. Res. Comm., 161, 851-58, 1989. Cells are plated at about 8 x 10³ cells per well in 12 well plates in DMEM supplemented with 10% calf serum, 2 mM glutamine, and antibiotics. Conditioned medium from transiently transfected 293 cells 72 hours post-transfection is added and cell number determined after 5 days.

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EXAMPLE 7

Assay for VEGF-R Antagonist

Assay reactions are set up essentially as described in Example 6, except that a compound to be tested for VEGF-R antagonist activity is included at the step of adding conditioned medium to bovine capillary endothelial cells. Multiple assays can be set up in which a constant amount of conditioned medium is incubated with varying amounts of test compound, for example from about 10 ng/ml to about 100 ug/ml.

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For conducting the cell growth assay, bovine capillary endothelial cells are maintained in DMEM containing 20% calf serum according to the method of Ferrara, Biochem. Biophys. Res. Comm., 161, 851-58, 1989. Cells are plated at about 8 x 10³ cells per well in 12 well plates in DMEM supplemented with 10% calf serum, 2 mM glutamine, and antibiotics.

Conditioned medium from transiently transfected 293 cells 72 hours post-transfection is added and cell number determined after 5 days.

WE CLAIM:

1. A substantially pure protein having the amino acid sequence designated herein as SEQ ID NO: 2.

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- 2. An isolated nucleic acid encoding the protein of Claim 1.
- 3. An isolated nucleic acid as in claim 2 wherein said nucleic acid is:

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- a) SEQ ID NO:1, or
- b) residues 276 through 1310 of SEQ ID NO:1.
- 4. An isolated nucleic acid that hybridizes to a nucleic acid defined by residues 276 through 1310 of SEQ ID NO:1 under high stringency conditions, said nucleic acid encoding a protein having angiogenic activity in vivo or in vitro.
- 5. A vector comprising an isolated nucleic acid defined 20 by residues 276 through 1310 of SEQ ID NO:1.
 - 6. A vector, as in Claim 5, wherein said isolated nucleic acid compound is operably-linked to a promoter sequence.

- 7. A host cell containing a vector of Claim 5.
- 8. A host cell containing a vector of Claim 6.
- 9. A method for constructing a recombinant host cell having the potential to express SEQ ID NO:2, said method

comprising introducing into said host cell by any suitable means a vector of Claim 6.

- 10. A method for expressing SEQ ID NO:2 in a recombinant host cell of Claim 9, said method comprising culturing said recombinant host cell under conditions—suitable-for-gene-expression.
- 11. A method for identifying compounds that bind a 10 protein identified herein as SEQ ID NO:2, comprising the steps of:
 - a) admixing a substantially purified preparation of a protein comprising SEQ ID NO:2 with a test compound; and
- b) monitoring by any suitable means a binding interaction between said protein and said compound.
- 12. A nucleic acid molecule comprising a fragment of SEQ ID NO:1, wherein said fragment consists of at least 14 contiguous base pairs therefrom.
 - 13. An antibody that selectively binds to a protein identified herein as SEQ ID NO:2.
- 25 14. A pharmaceutical formulation comprising as an active ingredient VEGF-R protein, associated with one or more pharmaceutically acceptable carriers, excipients, or diluents thereof.
- 15. A substantially pure protein as in Claim 1 wherein said protein is a dimer of SEQ ID NO:2.

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- 16. A pharmaceutical formulation comprising as an active ingredient an antagonist of an VEGF-R, associated with one or more pharmaceutically acceptable carriers, excipients, or diluents thereof.
- 17. A method for inhibiting tissue growth by administration of a therapeutically effective amount of an VEGF-R antagonist.

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- 18. A method for stimulating tissue growth by administration of an effective amount of an VEGF-R protein.
- 19. A method for identifying an VEGF-R antagonist comprising the steps of:
 - a) admixing a substantially purified preparation of VEGF-R with a test compound; and
 - b) monitoring, by any suitable means, an inhibition in the biological activity of VEGF-R.

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- 20. A method for stimulating angiogenesis comprising the step of administering to a mammal in need thereof a therapeutically effective amount of VEGF-R.
- 21. A method for treating coronary artery blockage comprising administering to a mammal in need thereof an effective amount of VEGF-R.
 - 22. An analog of SEQ ID NO:2.

- 23. A functional fragment of SEQ ID NO:2.
- 24. A substantially pure protein comprising the amino acid sequence selected from the group consisting of:
 - a. residues 16 through 345 of SEQ ID NO:2;
 - b. residues 23 through 345 of SEQ ID NO:2.

PCT/US99/01574 WO 99/37671

SEQUENCE LISTING

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Cys Val Cys Arg Gly Ser Thr Gly Gly 340 345

International application No. PCT/US99/01574

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A. CLASSIFICATION OF SUBJECT MATTER IPC(6) :Please See Extra Sheet.							
US CL : 530/350; 536/23.1; 435/7.1, 320.1, 252.3, 69.1; 424/277.1							
According to International Patent Classification (IPC) or to both national classification and IPC							
B. FIELDS SEARCHED							
Minimum documentation searched (classification system follows)							
U.S. : 530/350; 536/23.1; 435/7.1, 320.1, 252.3, 69.1; 42	24/277.1						
Documentation searched other than minimum documentation to	the extent that such documents are included in the fields searched						
	(name of data base and, where practicable, search terms used)						
BIOSIS, CANCERLIT, CAPLUS, EMBASE, IFIPAT, LIFE search terms: VEGF receptor, extracellular, fragment	SCI. MEDLINE, SCISEARCH, USPATFULL,GENBANK						
C. DOCUMENTS CONSIDERED TO BE RELEVANT							
Category* Citation of document, with indication, where	appropriate, of the relevant passages Relevant to claim No.						
X Database BIOSIS, No. 1998:123343,	LIN, P. et al. Inhibition of 14						
tumor growth by targeting tumor	endothelium using a soluble						
vascular endothelial growth factor							
Differentiation. January 1998, Vol. 49-58, see the abstract.	9, No. 1, pages 49-58, pages						
49-38, see the abstract.							
X Database BIOSIS, No. 1998:3	31506, TANAKA et al. 14						
Characterization of the extracellular	domain in vasular endothelial Cancer Res. September 1997,						
growth factor receptor-1. Japanese J							
Vol. 88, No. 9, pages 867-876, see	especially the abstract.						
į							
X Further documents are listed in the continuation of Box	C. See patent family annex.						
Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand						
"A" document defining the general state of the art which is not considered to be of particular relevance	the principle or theory underlying the invention						
earlier document published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step						
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other	when the document is taken alone						
special reason (as specified) *O* document referring to an oral disclosure, use, exhibition or other	Y document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is						
means	combined with one or more other such documents, such combination being obvious to a person skilled in the art						
the priority date claimed	*& document member of the same patent family						
Date of the actual completion of the international search	Date of mailing of the international search report						
06 MAY 1999	19 MAY 1999						
Name and mailing address of the ISA/US	Authorized officer						
Commissioner of Patents and Trademarks Box PCT	SUSAN UNGAR Tylenhour No. (703) 308 0196						
Washington, D.C. 20231 Facsimile No. (703) 305-3230	Telephone No. (703) 308-0196						

International application No.
PCT/US99/01574

	PCT/US99/015	/4
C (Continua	tion). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim N
x	Database BIOSIS, No. 1997:221744, BARLEON et al. Mapping of the sites for ligand binding and receptor dimerization at the extracellular domain of the vascular endothelial growth factor receptor FLT-1. J. Biol. Chem. 1997, Vol. 272, No. 16, pages 10382-10388, see especially the abstract.	14
	·	

Form PCT/ISA/210 (continuation of second sheet)(July 1992)*

International application No. PCT/US99/01574

Box 1 Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
Please See Extra Sheet.
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. X No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-12, 14, 15, 23, 24
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*

International application No. PCT/US99/01574

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

C07K 14/00; C07H 21/04; G01N 33/53; C12N 15/63, 15/85; A61K 35/12

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s)1-12, 18, 23, 24, drawn to a substantially pure VEGF receptor protein with amino acid sequence SEQ ID NO:2, nucleic acid encoding the sequence, a method of making the protein encoded by the nucleic acid sequence and a method of using the protein.

Group II, claim(s) 13, drawn to an antibody that binds to SEQ ID NO:2.

Group III, claim(s) 16, drawn to an antagonist of VEGF receptor.

Group IV, claim(s)17, drawn to a method for inhibiting tissue growth by administering a VEGF-R antagonist.

Group V, claim(s) 18, drawn to a method for stimulating tissue growth by administering VEGF receptor protein.

Group VI, claim(s) 19, drawn to a method for identifying a VEGF-receptor antagonist.

Group VII, claim(s) 20, drawn to a method for stimulating angiogenesis comprising administering VEGF-receptor. Group VIII, claim(s) 21, drawn to a method for treating coronary artery blockage comprising administering VEGF-

Group IX, claim(s) 22, drawn to an analog of SEQ ID NO:2.

The inventions listed as Groups I-VIII do not relate to a single inventive concept under PCT Rule 13.1 because, under Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

The application shall relate to one invention only or to a group of inventions so linked as to form a single general inventive concept. if multiple products, processes of manufacture or uses are claimed, unity of invention is determined by 37 CRF 1.475(b) which defines the combinations of categories held to have unity of invention. See PCT article 17(30(a) and 37 CFR 1.475(b). After that all other products and processes will be broken out as separate inventions.

The special technical feature of Group I is considered to be the VEGF receptor protein with amino acid SEQ ID NO:2. The special technical feature of Group II is considered to be an antibody that binds to SEQ ID NO:2, not found in Group I

The special technical feature of Group III is considered to be an antagonist of VEGF receptor, not found in Group I. The special technical feature of Group IV is considered to be a method for inhibiting tissue growth by administering a VEGF-receptor antagonist, not found in Group I.

The special technical feature of Group V is considered to be a method for stimulating tissue growth by administering a VEGF-receptor protein, not found in Group I.

The special technical feature of Group VI is considered to be a method for identifying a VEGF-receptor antagonist, not found in Group I.

The special technical feature of Group VII is considered to be a method for stimulating angiogenesis, not found in Group I.

The special technical feature of Group VIII is considered to be a method for treating coronary artery blockage, not found in Group I.

The special technical feature of Group IX is considered to be an analog of SEQ ID NO:2, not found in Group I.

Accordingly Groups I-IX are not so linked by the same or a corresponding special technical feature as to form a single general inventive concept.

Form PCT/ISA/210 (extra sheet)(July 1992)*